Short term maternal psychological stress in the post-conception period in ewes affects fetal growth and gestation length.

Short title: Post-conception stress and fetal development.

Jennifer Smith¹, Drewe Ferguson¹, Guillermo Jauregui², Martín Panarace², Mariano Medina², Sigrid Lehnert³, Jonathan R. Hill¹

¹CSIRO Livestock Industries, FD McMaster Laboratory, Armidale, Australia
²Goyaie S.A.A.C.I y F Biotecnología Animal, Carmen de Areco, Buenos Aires, Argentina and ³CSIRO Livestock Industries Queensland Biotechnology Precinct, St Lucia, Australia.

Correspondence should be addressed to: Jonathan Hill, email Jon.hill@csiro.au
ABSTRACT

Fetal development can be influenced by maternal environment in the periconceptional period. This study investigated the effect of maternal feed intake and psychological stress within the first 6 days after conception, on embryo development and fetal growth. Superovulated ewes (n=40) were artificially inseminated with semen from one ram. Ewes were then divided into four groups (n=10). Group 1, (Control), was fed at maintenance level, group 2 (High) at 2 x maintenance and group 3 (Low) at 0.5 x maintenance on Days 2-6 after conception. Group 4 (Stress) was fed at maintenance level and then an intense physical and psychological stress challenge was applied for 1 hour only on Days 2 and 3 after conception. Embryos were recovered at Day 6. A total of 113 transferable grade embryos were transferred singly into synchronized untreated recipients while the remaining embryos (n=165) were fixed and stained for cell counts. Post conception maternal stress or feed intake did not alter the cell count or grade of Day 6 embryos. Fetuses from the stress group had longer crown rump lengths at Day 30 and longer femur length at Day 58. Fetuses from the stressed and high feed groups had greater abdominal circumferences at Day 85. Subsequent birth weights were not significantly different. Ewes carrying lambs from the stress treatment had shorter gestation lengths. These results show that short term perturbations of the post-conception maternal environment, have measurable effects on fetal development and gestation length.
INTRODUCTION

The maternal environment in the periconceptional period can impact upon fetal or placental growth, gestation length and post natal health (Armitage et al., 2004, Fleming et al., 2004, McMillen & Robinson, 2005, Oliver et al., 2007). This has important implications for human health. In commercial animal production, the notion that the animal’s maximum production potential and health is affected by conditions during pregnancy creates an imperative to identify the critical time periods and mechanisms for fetal programming.

Nutrient restricted diets around the time of conception in sheep have been shown to affect fetal development and physiology. Diet restriction for 2-3 weeks prior to mating resulted in altered embryo morphology and increased embryonic trophectoderm cell numbers (Abecia et al., 1997, Nolan et al., 1998). Gestation length was reduced by a week in ewes as a result of severe nutrient restriction from one month prior to conception through to month 2 of gestation (Bloomfield et al., 2003). More moderate feed restriction from 60 days prior to through to 7 days after conception resulted in earlier hypothalamo-pituitary adrenal axis activation in twin but not singleton pregnancies (Edwards & McMillen, 2002). Nutrient excess, via high feed intake from Day 5-55 of gestation, reduced fetal cotyledon mass and lamb birth weight (Wallace et al., 2003). In gilts fed 2 x maintenance from the day after conception for 2 weeks, higher embryonic losses were associated with lower maternal serum progesterone (Jindal et al., 1996).

Events during the week after conception have been shown to influence subsequent development, which demonstrates the sensitivity of early embryos to environmental cues Progesterone supplementation during the first few days of
pregnancy results in a significant increase in crown rump length in ovine fetuses (Garrett et al., 1988, Kleemann et al., 1994). Increased birth weights result from in vitro culture of Day 0-6 ovine embryos in the presence of serum which supplies an excess of energy substrates and growth factors (Walker et al., 1996).

These effects led us to target Days 2-6 post-conception as the period to alter the in vivo maternal environment during the time when embryonic development is progressing from cleavage stage through to blastocyst formation. Altered feed intake and imposition of a short term psychological stress were used as periconceptional treatments as these are commonly encountered aspects of animal production and human society. We hypothesized that high cortisol levels induced by stress would increase energy substrate availability in the oviducts and alter early embryonic development. In dairy cattle, stress around the time of conception results in depressed pregnancy rates, and therefore practitioners of assisted reproductive technologies usually recommend strategies to reduce stress during this critical period (Dobson et al., 2001).

The aim of the present study was to investigate whether fetal growth and development can be altered by maternal stress and feed intake changes applied in the first week after conception. As in vitro cultured embryos are highly sensitive to treatments during this first week, we hypothesized that in vivo embryos may be similarly affected by altering the maternal environment during this period. Measurable effects would illustrate the sensitivity of the developing embryo to perturbations in maternal environment around the time of conception and provide a model to study mechanisms and subsequent impacts on fetal development.

MATERIALS AND METHODS
**Ewe selection and preparation**

All procedures were carried out in accordance with the Australian Code of Practice for the Use of Animals for Scientific Purposes and were approved by the CSIRO Livestock Industries Animal Ethics Committee. Body weight (BWT) and body condition score (BCS scale 1 – 5) of 160 Merino ewes were recorded prior to the allocation of ewes to donor or recipient groups. These measures were then repeated prior to artificial insemination (AI) and several times throughout pregnancy. Ewes were allowed to adjust to the pellet ration and animal house conditions for 1 week prior to estrus synchronization and allocation to treatment groups. The ration was lucerne based commercial sheep pellets (8 MJ ME/kg, 15.5% Crude Protein). Forty ewes were selected as embryo transfer (ET) donors with mean BWT of 40 kg and BCS of 3.0, while the remaining 120 ewes were allocated to be recipients.

Donor ewes were superovulated and inseminated as 4 separate day groups (10 per group). On Day 2 after AI they were separated and allocated to the following treatment groups: (i) Control - maintenance Day 2-6 after conception; (ii) High feed – 2 x maintenance Day 2-6 after conception; (iii) Low feed - 0.5 x maintenance Day 2-6 after conception and (iv) Stress - maintenance Day 2-6 after conception + stress on Days 2 and 3.

Ewes were distributed from each treatment group across the 4 days. Thus on each of the 4 separate days, embryos were recovered from 10 ewes during the morning then on the same afternoon embryos were either transferred into at least 25 recipients or retained for cell counts.

**Feed intake and stress treatments**
In the week prior to, and after AI, donors were fed individually in single pens. All ewes were fed at maintenance (0.8 kg/d) in the week up to, and the day after AI. The experimental treatments commenced on day 2 post-AI. The donor ewes were returned to pasture after ET on day 6.

The stress treatment comprised a combination of physical and psychological stressors including transport from the animal pens to nearby shearing shed/sheep yards; shearing (half belly wool shorn off on each day); exposure to a sheep dog for 1 hr duration; 10 min isolation from other sheep followed by movement back to their pens with the assistance of a dog. Blood samples (6 ml K$_2$-EDTA vacutainers) for measurement of plasma cortisol were collected by jugular venipuncture from Control and Stress group ewes on the stress treatment days. Blood samples were collected immediately prior to commencement of the stress period ($t_0$); approximately midway through the stress period ($t_1$); and 15 min after cessation of the stress period ($t_2$). Samples were collected on both days 2 and 3. The control group ewes remained in their pens and were bled at similar times to the stress group ewes. Plasma cortisol was determined using a commercial radioimmunoassay (Spectria Cortisol RIA, Orion Diagnostica, Espoo, Finland), adapted and validated for ovine plasma (Paull et al., 2007).

Superovulation, insemination and embryo transfer

The estrous cycles of donor and recipient ewes were synchronized using progesterone implants (CIDR-g, Carter-Holt) for 14 days. Superovulation of donor ewes was achieved by a twice daily decreasing treatment of follicle stimulating hormone (Folltropin, Bioniche) for 4 days concluding 24 hr after CIDR removal. Equine chorionic gonadotropin (eCG 200 IU; Pregnecol, Bioniche) was administered
i.m. the day before CIDR removal. Laparoscopic insemination was carried out approximately 39 hr after CIDR removal using freshly collected semen from a single sire (Day 0). On Day 6, embryos were recovered from the uterus under general anaesthesia using ViGro Complete Flush Solution (Bioniche, Belleville, Canada). Recovered embryos were washed in ViGro then held in HEPES buffered SOF embryo holding medium (Cook Embryo Holding Medium, Brisbane, QLD) at room temperature (24°C) to enable grading and sorting. Embryos for transfer were held for 2-4 hours prior to transfer into recipient ewes. Transfer of embryos into the uterus was via laparoscopic assistance under heavy sedation (3 mg Xylazine i.m.). Embryos were randomly transferred into recipient ewes.

**Embryo grading and cell counts**

Embryo morphology was evaluated under a stereomicroscope for grading according to developmental stage (morula through to expanded blastocyst) and quality (1 to 4). Embryos were classified as transferable quality when they achieved a Grade 1 or 2 and they were at the appropriate stage of development for Day 6 after fertilization (compact morula or blastocyst). Embryos of transferable grade were randomly assigned to be either transferred into a recipient (single embryo per recipient) or fixed and stained for cell counting.

Total cell counts were established by equilibration of embryos in 0.5 mg/ml Hoechst 33342 for 30 min. Compact morula, morula and degenerate embryos were then visualized under UV light using a blue/green filter. Blastocysts were differentially stained to show inner cell mass (ICM) and trophectoderm cells. Differential staining was accomplished by disrupting membrane integrity on the surface trophectoderm cells with detergent (TritonX-100) together with staining by
propidium iodide and Hoechst 33342 (Thouas et al., 2001). This permitted penetration of propidium iodide into the trophectoderm cells but not the ICM cells. As all cells were stained with Hoechst 33342, the ICM stained blue and the trophectoderm red. All evaluations were performed without knowledge of treatment groups.

Ultrasonographic evaluation of fetuses

Pregnancy was determined by ultrasonography on day 30 of gestation. Thereafter fetal heart rate (beats/min) and growth measures were recorded by ultrasound scanning on days 30, 44, 58, 85, 114 and 142 of gestation. The operator was blind to treatment groups. On day 30 and 44, fetal assessment was performed transrectally and fetal size was assessed as crown-rump length (mm). Thereafter, scanning was transabdominal and fetal crown rump was replaced by femur length (mm) and abdominal circumference (mm). Each parameter was measured at least twice and the mean recorded.

Parturition

Lambing took place in indoor pens under constant supervision during the day and every 6 hrs at night. This allowed recovery of the majority of placentas and for the time of parturition to be recorded within 6 hrs of birth. Lamb records included birth date, sex, weight, surrogate dam (embryo recipient) and genetic dam (embryo donor). Intact placentas were weighed then the cotyledons were counted, dissected off the membranes and weighed separately from the remaining placenta.

Statistical analysis
Data were analysed using ASReml (Gilmour et al., 2002). Analysis of variance was conducted to test for the main effects of maternal feed intake (control, high, low), stress (stress, unstressed), day of ET (4 days) and where appropriate, fetal sex on embryonic cell count, pregnancy rate, fetal heart rate and growth, birth weight, and placental measurements. Where appropriate, maternal bodyweights were fitted as covariates and first order interactions were included. Donor was fitted as a random effect to the cell count data. Stress and feed were retained in the final model and non-significant main effects and interactions iteratively removed. Data are presented as ASReml predicted means ± standard errors of the means (S.E.M.). All traits examined were tested for statistical normality and no data transformations were required.

RESULTS

**Plasma Cortisol**

Changes in plasma cortisol were determined using a repeat measures analysis. Cortisol level at t₀ (basal level) was fitted as a covariate and was not different for Control and Stress groups. Cortisol level of the Stress group were elevated at t₁, and, although declining by t₂, remained significantly higher than Controls (P<0.001) (Figure 1). Neither day of ET (4 levels) nor bleed day (days 2 and 3 after AI) were significant effects on cortisol level.

**Embryo morphology and cell counts**

Surgical flushing of the 40 superovulated ewes yielded 343 embryos with a fertilization rate of 97.1%. Of the collected embryos, 82% of embryos were graded as transferable (Table 1). Of the transferable embryos, 113 were surgically transferred
singly into recipient ewes; the remainder were fixed and stained for cell counting. None of the treatments improved the proportion of transferable embryos.

Adjusted mean total cell counts of all recovered embryos and of the selected transferable quality embryos were not significantly affected by treatment (Table 1). There was a significant day effect on total cell count and inner cell mass/trophectoderm ratio where the highest total cell counts were observed on the last day (day 4) of ET.

**Pregnancy rates**

At Day 30 after AI, the overall pregnancy rate was 61%. Pregnancy rates differed significantly between the treatments, where the lowest and the highest rates were observed for the High feed (44%) and Stress groups (80%), respectively (p<0.05; Figure 2). Other treatment group contrasts were not significant. Day of transfer and recipient body weights were not significant effects which indicated recipient animals were evenly spread between groups and days.

**Fetal Development**

**Stress.** At day 30, crown rump lengths of fetuses from Stress ewes were 4.4% longer than controls (P<0.05; Table 2). This difference approached significance at Day 44 (P=0.10) then at Day 58 femur length was longer (P<0.05). At Day 85, abdominal circumference was larger in the Stress group compared to controls (P<0.01). A month later (Day 114), there was no difference between the treatment groups in femur length or abdominal circumference and this continued to Day 142. Throughout pregnancy there were no effects of Stress on fetal heart rate.

**Feed intake.** At days 58 and 85, fetuses from High feed ewes had larger abdominal circumferences than controls (P<0.05; Table 2). Heart rate at Day 44 was
elevated in both High and Low feed intake groups and again in the High feed group at Day 58.

Effect of Day. The day of ET had an effect on fetal growth up to day 44 (P<0.05 at day 30 and P<0.001 at day 44). Fetuses transferred on day 4 were significantly longer than those from other ET days (eg at Day 30, Day 4, 19.8 ± 1 vs Day 1, 17±1.2, Day 2, 17.8 ±0.7, Day 3, 17 ±0.7). This correlated with increased embryo cell number on Day 4. The strong effect of day of transfer in our experiment represented random environmental effects that persisted through pregnancy but were evened out by birth.

Parturition and gestation length

Gestation lengths for the Stress fetuses were shorter (1.3 days) than for Controls (P<0.05, Table 3). The effect of maternal feed intake approached significance (P=0.065) for cotyledon count where the count for the High feed group trended lower than the Control and Low feed groups. There were no significant treatment effects on placenta weight, or the weight of cotyledons as a proportion of total placenta weight. There was a significant interaction between treatment and day of ET for placenta weight and cotyledon proportion. Embryos from the Control and Stress groups transferred on the first 3 days were similar, but among the embryos transferred on day 4, the Stress group resulted in fetuses with much larger (almost double) placenta weight and cotyledon proportion. Sex did not affect birth weight and there were no indications of treatment effects on birth weight (Table 3).

DISCUSSION

By focusing on the first week after conception, we have demonstrated that brief periods of maternal stress and elevated feed intake induced measurable changes
in fetal development. One hour of physical and psychological stress applied on two consecutive days during the first week following conception significantly increased fetal size during the first half of pregnancy and reduced gestation length. Increasing feed intake from Day 2-5 after conception was found to increase fetal size at Day 85 of gestation.

Fetuses with accelerated growth have resulted from exposure of embryos to a uterine environment with high substrate availability. For example, progesterone supplementation of ewes during the first 3 days after conception, as well as high maternal nutrient availability in the preimplantation period resulted in increased fetal growth rates (Kleemann et al., 1994, McEvoy et al., 1997). The brief stress applied to ewes in the current experiment could also have resulted in increased substrate availability to the embryo. The elevated serum cortisol levels observed in our study should have lead to mobilization of energy stores and results in an increase in the levels of glucose, free fatty acids and amino acids (Sapolsky, 2002). The increase in crown-rump length observed in the Stress group at Day 30 could therefore be the result of increased nutrient availability, although we did not measure this in the ewes during the treatment period. Potential mechanisms for this effect include the impact of glucose concentration on the expression of glucose transporter molecules in the preimplantation embryo as well as other membrane transporters (Pantaleon & Kaye, 1998, Jansen et al., 2006). Subtle alterations of the biochemical environment of the preimplantation embryo could also exert direct effects on the chromatin modification processes that take place during this period of development.

Some stressors during gestation can alter human birth weights and there is an increased risk of preterm birth in women with high levels of long term stress (Dole et al., 1990).
Prenatal stress has been studied in Rhesus monkeys but only during mid gestation (Schneider, 1992). To model human gestational stress, studies in rodents have typically focused on mid to late gestation stressors with an outcome of behavioral and psychological changes in offspring (Kofman, 2002). In one rodent study where early gestational stress (Day 6) was investigated, the stressed pregnant rats gave birth to fewer females and the pups were larger by 4 weeks after birth (Mueller & Bale, 2006).

The short term psychological stress, applied here in the early post-conception period, resulted in a significant reduction in gestation length. The reduction by 1 day was small compared to the 7 day reduction observed by Bloomfield et al. (Bloomfield et al., 2003). In that study, the ewes were underfed from Day -60 to Day +30 of gestation, and the nutritional deprivation resulted in a 15% reduction in ewe bodyweight. The mechanism by which gestation length can be altered by an early gestational treatment is thought to be through accelerated development of the hypothalamic-pituitary adrenal axis (Edwards & McMillen, 2002). This in turn, may lead to a shift in the timing of the fetal cortisol peak that precedes parturition. It is unclear whether the effect of maternal undernutrition on the early embryo is exerted through exposure to an altered metabolic environment or to elevated glucocorticoid concentrations in response to nutritional stress. The results from the present study appear to indicate that short term exposure of the sheep preimplantation embryo to elevated cortisol (and adrenaline) levels led to subtle reprogramming of the hypothalamic-pituitary axis.

Embryos from high feed intake (2 x maintenance) ewes developed into fetuses with greater abdominal circumferences at Day 85 (P<0.05). Cotyledon counts tended to be lower (p=0.065) in the high feed intake than maintenance or low feed groups.
which is in agreement with Wallace et al., 1999, who found a small significant reduction on cotyledon count at term in ewes fed a high feed intake for the first 50 days of gestation. However, high maternal feed intake does not consistently affect placentome numbers nor fetal dimensions (Quigley et al., 2005). We did not find any effect of periconceptional undernutrition on fetal or placental parameters at term similar to that reported by MacLaughlin et al., 2005.

Possible mechanisms of the increased early fetal growth observed in the Stress and the High nutrition groups include altered imprinted gene expression, activation of universal stress response pathways in the embryo, or interference with embryo-maternal signaling. The postconception period in sheep embryos is characterized by a prolonged period of active chromatin demethylation and re-methylation (Beaujean et al., 2004). This may partly explain the particular sensitivity of sheep embryos to IVF culture media (Walker et al., 1996) and to changes of the uterine environment in vivo, such as those resulting from progesterone supplementation (Kleemann et al., 1994).

We did not find any effect of treatment on embryo stage or morphological grade that could not be explained by the effect of day of ET. This contrasted with other results where high nutrition accelerated development towards the expanded blastocyst stage (Abecia et al., 1997). Our results showed a high proportion of embryos were graded as good quality (control 88%, Low 88%, High 81%, Stress 73%), whereas embryos from ewes offered high/low diets from Day -10 through Day +4 were graded lower quality than controls (Lozano et al., 2003).

Pregnancy rates were affected by the treatment of the donor ewes. This was anticipated, however, the significantly higher pregnancy rate in the Stress group compared to the High feed group was not expected. Parr et al., 1987 showed that high
feed intake on Days 2-14 after mating resulted in reduced pregnancy rates in ewes and this lowered pregnancy rate was reversed by progesterone supplementation.

This study provides evidence of fetal responses to mild changes in maternal environmental stress and nutrition and provides further evidence of fetal responses to altered maternal environment. It sets the basis from which to explore the effects of psychological stress in its various manifestations on fetal development. This has implications for both livestock productivity and human health. We used a short term stress – just 1 hour – and in future experiments we propose to investigate longer periods of stress which are regularly encountered by livestock (eg storms, feed deprivation, handling stresses). In modern society, women during early pregnancy are also exposed to stressful environments and the sheep model developed here may produce relevant information to correlate with human situations.

ACKNOWLEDGEMENTS

Supported by funding from Goyaike S.A.A.C.I y F Biotecnología Animal, Buenos Aires, Argentina and CSIRO Livestock Industries, Brisbane, Australia. L. Piper provided valuable statistical advice, J. Lea performed the cortisol assays, S. Atkinson, R. Davey and K. Hutton assisted with the embryo transfer, and D. Niemeyer and S. Belson assisted with sampling and feeding, all of which was greatly appreciated.

REFERENCES


Figure Legends

**Figure 1.** The severity of the stress event applied to ewes on 2 consecutive days (Day 2,3 after fertilization) was illustrated by significant elevations in cortisol levels both during (Time 1) and after (Time 2) application of stress. Time 0 represents basal cortisol levels in treated and untreated groups, Time 1 illustrates cortisol values from samples obtained during the period of stress. Time 2 is one hour later, and shows that cortisol remained elevated 15 minutes after ewes were returned to their pens, alongside control animals (values with different superscripts are significantly different at p<0.05).

**Figure 2.** Pregnancy rates at Day 30 were significantly lower for embryos exposed to a high feed environment compared to embryos exposed to stress after conception (values with different superscripts are significantly different at p<0.05). Contrary to our hypothesis, pregnancy rates from stressed embryos were not lower than those from controls.
The severity of the stress event applied to ewes on 2 consecutive days (Day 2,3 after fertilization) was illustrated by significant elevations in cortisol levels both during (Time 1) and after (Time 2) application of stress. Time 0 represents basal cortisol levels in treated and untreated groups, Time 1 illustrates cortisol values from samples obtained during the period of stress. Time 2 is one hour later, and shows that cortisol remained elevated 15 minutes after ewes were returned to their pens, alongside control animals (values with different superscripts are significantly different at p<0.05).
Pregnancy rates at Day 30 were significantly lower for embryos exposed to a high feed environment compared to embryos exposed to stress after conception (values with different superscripts are significantly different at p<0.05). Contrary to our hypothesis, pregnancy rates from stressed embryos were not lower than those from controls.
Table 1. Number of embryos collected and embryo cell counts from superovulated donor ewes (actual embryo counts and adjusted cell count means ± s.e.)

<table>
<thead>
<tr>
<th>Embryo analysis</th>
<th>Control</th>
<th>High feed</th>
<th>Low feed</th>
<th>Stress</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number embryos recovered</td>
<td>80</td>
<td>96</td>
<td>68</td>
<td>99</td>
<td>343</td>
</tr>
<tr>
<td>Raw mean transferable grade (% of total)#</td>
<td>88%</td>
<td>81%</td>
<td>88%</td>
<td>73%</td>
<td>82%</td>
</tr>
<tr>
<td>Adjusted mean transferable grade embryos$</td>
<td>74% ± 9</td>
<td>76% ± 9</td>
<td>88% ± 9</td>
<td>78% ± 9</td>
<td></td>
</tr>
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</table>

<table>
<thead>
<tr>
<th>Cell counts</th>
<th>All embryos</th>
<th>Transferable grade embryos</th>
<th>Inner cell mass: Trophoderm ratio</th>
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<tbody>
<tr>
<td>Number embryos transferred</td>
<td>37.2 ± 5.8</td>
<td>44.8 ± 4.7</td>
<td>36.7 ± 0.05</td>
</tr>
<tr>
<td>Number embryos transferred</td>
<td>46.0 ± 5.4</td>
<td>49.2 ± 4.4</td>
<td>32.7 ± 0.05</td>
</tr>
<tr>
<td>Number embryos transferred</td>
<td>48.8 ± 5.5</td>
<td>52.1 ± 4.4</td>
<td>35.5 ± 0.11</td>
</tr>
<tr>
<td>Number embryos transferred</td>
<td>37.6 ± 5.5</td>
<td>46.4 ± 4.7</td>
<td>36.2 ± 0.09</td>
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<table>
<thead>
<tr>
<th>Embryo survival</th>
<th>Number embryos transferred</th>
<th>Ewes pregnant at D 30</th>
</tr>
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<tbody>
<tr>
<td>Number embryos transferred</td>
<td>29</td>
<td>68% ± 9</td>
</tr>
<tr>
<td>Number embryos transferred</td>
<td>34</td>
<td>44% ± 8</td>
</tr>
<tr>
<td>Number embryos transferred</td>
<td>25</td>
<td>55% ± 11</td>
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<tr>
<td>Number embryos transferred</td>
<td>25</td>
<td>80% ± 10*</td>
</tr>
</tbody>
</table>

# raw % transferable grade of all embryos collected
$ adjusted mean ± SE (donor was included as a random effect in the statistical model)
* p<0.05
Table 2. Assessment of treatment effects during pregnancy by ultrasonographic measurements of fetal parameters (mean and s.e.).

<table>
<thead>
<tr>
<th>Fetus Age</th>
<th>Parameter</th>
<th>Control (n=19)</th>
<th>High feed (n=14)</th>
<th>Low feed (n=13)</th>
<th>Stress (n=20)</th>
<th>Stress effect</th>
<th>Feed effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>30</td>
<td>Heart rate</td>
<td>194.5 ± 2.6</td>
<td>195.5 ± 2.8</td>
<td>194.0 ± 3.0</td>
<td>196.7 ± 2.5</td>
<td>ns</td>
<td>ns</td>
</tr>
<tr>
<td></td>
<td>Length (crown rump)</td>
<td>17.6 ± 0.3</td>
<td>17.5 ± 0.3</td>
<td>17.9 ± 0.3</td>
<td>18.4 ± 0.3</td>
<td>P&lt;0.05</td>
<td>ns</td>
</tr>
<tr>
<td>44</td>
<td>Heart rate</td>
<td>212.3 ± 2.9</td>
<td>219.8 ± 3.1</td>
<td>222.1 ± 3.3</td>
<td>218.2 ± 2.8</td>
<td>ns</td>
<td>P&lt;0.05</td>
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<td></td>
<td>Length (crown rump)</td>
<td>45.8 ± 0.7</td>
<td>45.9 ± 0.8</td>
<td>46.1 ± 0.8</td>
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<tr>
<td>58</td>
<td>Heart rate</td>
<td>216.3 ± 2.9</td>
<td>222.2 ± 3.2</td>
<td>219.4 ± 3.4</td>
<td>217.3 ± 2.9</td>
<td>ns</td>
<td>P&lt;0.05</td>
</tr>
<tr>
<td></td>
<td>Femur length</td>
<td>8.5 ± 0.2</td>
<td>8.7 ± 0.2</td>
<td>8.5 ± 0.2</td>
<td>8.8 ± 0.2</td>
<td>P&lt;0.05</td>
<td>ns</td>
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<tr>
<td></td>
<td>Abdominal circumf</td>
<td>88.1 ± 1.0</td>
<td>91.1 ± 1.0</td>
<td>88.7 ± 1.1</td>
<td>90.0 ± 1.1</td>
<td>ns</td>
<td>P&lt;0.05</td>
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<tr>
<td>85</td>
<td>Heart rate</td>
<td>192.7 ± 4.1</td>
<td>195.1 ± 4.4</td>
<td>192.1 ± 4.7</td>
<td>191.8 ± 4.0</td>
<td>ns</td>
<td>ns</td>
</tr>
<tr>
<td></td>
<td>Femur length</td>
<td>24.6 ± 0.3</td>
<td>24.6 ± 0.3</td>
<td>24.0 ± 0.4</td>
<td>24.8 ± 0.3</td>
<td>ns</td>
<td>P= 0.06</td>
</tr>
<tr>
<td></td>
<td>Abdominal circumf</td>
<td>165.1 ± 2.2</td>
<td>170.5 ± 2.3</td>
<td>164.7 ± 2.5</td>
<td>170.8 ± 2.1</td>
<td>P&lt;0.01</td>
<td>P&lt;0.05</td>
</tr>
<tr>
<td>114</td>
<td>Heart rate</td>
<td>167.8 ± 3.4</td>
<td>168.9 ± 3.7</td>
<td>170.8 ± 4.0</td>
<td>166.1 ± 3.4</td>
<td>ns</td>
<td>ns</td>
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<tr>
<td></td>
<td>Femur length</td>
<td>51.1 ± 0.5</td>
<td>50.9 ± 0.5</td>
<td>50.6 ± 0.5</td>
<td>51.0 ± 0.5</td>
<td>ns</td>
<td>ns</td>
</tr>
<tr>
<td></td>
<td>Abdominal circumf</td>
<td>269.5 ± 3.6</td>
<td>269.1 ± 3.9</td>
<td>267.2 ± 4.2</td>
<td>266.1 ± 3.6</td>
<td>ns</td>
<td>ns</td>
</tr>
<tr>
<td>142</td>
<td>Heart rate</td>
<td>132.7 ± 4.4</td>
<td>131.7 ± 4.8</td>
<td>132.9 ± 5.4</td>
<td>130.4 ± 4.4</td>
<td>ns</td>
<td>ns</td>
</tr>
<tr>
<td></td>
<td>Femur length</td>
<td>78.1 ± 0.5</td>
<td>78.2 ± 0.5</td>
<td>77.5 ± 0.6</td>
<td>78.4 ± 0.5</td>
<td>ns</td>
<td>ns</td>
</tr>
<tr>
<td></td>
<td>Abdominal circumf</td>
<td>324.5 ± 4.8</td>
<td>326.6 ± 5.2</td>
<td>325.9 ± 5.9</td>
<td>325.7 ± 4.8</td>
<td>ns</td>
<td>ns</td>
</tr>
</tbody>
</table>

# Fetal length and circumference measurements are in mm, heart rate in beats per minute.
ns: non-significant (p > 0.05)
Table 3. Treatment effects on birth weight and placental characteristics (mean and s.e.)

<table>
<thead>
<tr>
<th></th>
<th>Control (n=19, 14)#</th>
<th>High Feed (n=14, 12)#</th>
<th>Low Feed (n=11, 10)#</th>
<th>Stress (n=20, 16)#</th>
<th>Stress effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gestation length (days)</td>
<td>154.2 ± 0.5</td>
<td>153.3 ± 0.6</td>
<td>153.5 ± 0.6</td>
<td>152.9 ± 0.4</td>
<td>P &lt; 0.05</td>
</tr>
<tr>
<td>Placenta wt (g)</td>
<td>341.2 ± 33.9</td>
<td>325.9 ± 34.7</td>
<td>294.1 ± 37.7</td>
<td>346.3 ± 33.2</td>
<td>ns</td>
</tr>
<tr>
<td>Cotyledon number</td>
<td>67.1 ± 5.7</td>
<td>60.0 ± 6.0</td>
<td>62.7 ± 6.3</td>
<td>66.2 ± 5.5</td>
<td>ns</td>
</tr>
<tr>
<td>Ratio cotyledon:placenta weights</td>
<td>0.26 ± 0.03</td>
<td>0.26 ± 0.03</td>
<td>0.29 ± 0.03</td>
<td>0.29 ± 0.02</td>
<td>ns</td>
</tr>
<tr>
<td>Birth wt (kg)</td>
<td>4.3 ± 0.4</td>
<td>4.6 ± 0.4</td>
<td>4.2 ± 0.5</td>
<td>4.7 ± 0.3</td>
<td>ns</td>
</tr>
</tbody>
</table>

\#Number of observations (n) are for gestation length/birth weight and placental characteristics respectively