Superoxide dismutase activity, lipid peroxide production and corpus luteum steroidogenesis during natural luteolysis and regression induced by oestradiol deprivation of the ovary in pseudopregnant rabbits

J. S. Hesla*, T. Miyazaki†, L. M. Dasko, E. E. Wallach and
A. M. Dharmarajan

*Department of Gynecology and Obstetrics, The Johns Hopkins University School of Medicine,
Baltimore, MD 21205, USA*

Summary. The relationship of oxygen free radicals to corpus luteum function in rabbits was explored during various stages of pseudopregnancy, including natural and induced luteal regression. Induced luteolysis was achieved during mid-pseudopregnancy by removal of an oestradiol capsule placed at the onset of pseudopregnancy, which suppressed ovarian oestradiol production. Activity of manganese superoxide dismutase (Mn SOD) was significantly and positively correlated with ovarian progesterone production ($P < 0.01$) throughout pseudopregnancy and during natural regression. Oestradiol deprivation for 12, 24 or 72 h resulted in declines in Mn SOD activity and progesterone secretion, although Mn SOD rose and corpus luteum steroidogenesis was restored to normal when the capsule was replaced for 48 h before assessment, having been removed for 24 h. Lipid peroxide and progesterone concentrations were not correlated, although a significant rise in lipid peroxides in the luteal tissue was detected after deprivation of oestradiol for 72 h. Changes in progesterone production and Mn SOD activity were not associated with alterations in concentration of prostaglandin F metabolite. These data suggest that Mn SOD may be involved in regulating function of the corpus luteum during pseudopregnancy in rabbits and that oxygen free radicals may play a role in regression of corpus luteum in this species.

Keywords: corpus luteum; rabbit; superoxide dismutase; progesterone; lipid peroxide

Introduction

Although ovarian function in pseudopregnant rabbits has been well characterized (Dharmarajan *et al.*, 1989; Holt, 1989), the regulatory factors that influence growth, hormone production and regression of the corpus luteum remain to be clarified. Oestradiol is the primary luteotrophin in intact and hypophysectomized pseudopregnant rabbits; progesterone secretion declines dramatically when oestradiol support of the corpora lutea is withdrawn (Bill & Keyes, 1983; Dharmarajan *et al.*, 1991). The mechanism for this induced, premature regression of luteal function is not understood; recent data suggest that prostaglandins are not directly involved (Dharmarajan *et al.*, 1989), but there is increasing evidence from studies in rats that an accumulation of toxic metabolites of oxygen may be associated with luteolysis (Behrman & Preston, 1989; Behrman & Aten, 1991; Gatzuli *et al.*, 1991; Riley & Behrman, 1991; Sawada & Carlson, 1989, 1991).

*Reprint requests.

†Present address: Department of Obstetrics and Gynecology, Ogikubo Hospital, Suginami-ku, Tokyo 167, Japan.

An *in vivo* examination of the role of reactive oxygen species in cellular events is severely compromised by the extremely short half-lives of these metabolites. One indirect approach frequently used to characterize free-radical mechanisms in physiological processes is to measure the highly specific inhibitor of the superoxide anion, superoxide dismutase (SOD). Superoxide dismutase is thought to be induced by reactive oxygen species (Dryer *et al.*, 1980) and has been histochemically localized in rat ovary where it may play a role in regulating luteal function (Laloraya *et al.*, 1988).

Few data have been published concerning the ovarian action of superoxide radicals in species other than rat, although one recent study in rabbits clearly demonstrated a role for superoxide in the mechanical process of ovulation (Miyazaki *et al.*, 1991). The objectives of this investigation were to determine whether oxygen free radicals are involved in the regulation of the function of the corpus luteum in rabbits during pseudopregnancy and to assess the hypothesis that oestradiol influences luteal function by directly or indirectly promoting the production of free-radical scavengers. Corpus luteum SOD activity and lipid peroxide concentrations were measured and correlated with progesterone production at several points during the natural lifespan of the corpus luteum in pseudopregnancy. Because of its physiological role in the process of ovulation and corpus luteum regression in many animals, prostaglandin production was measured concurrently. The known requirement of oestradiol support for functional maintenance of the corpora lutea allowed for the development of an *in vivo* model to assess the effects of acute shifts in concentration of circulating oestradiol on production of free-radical scavengers during mid-pseudopregnancy.

Materials and Methods

Animals

Sexually mature New Zealand White female rabbits with a mean weight of 4.0 kg were used. The animals were caged individually for a minimum of 3 weeks under controlled light and temperature and given free access to water and Purina Rabbit Chow (Ralston-Purina Co., St Louis, MO, USA). Pseudopregnancy was induced by administration of human chorionic gonadotrophin via marginal ear vein (hCG, 100 iu; Pregnyl; Organon, Inc., West Orange, NJ, USA). The day of hCG injection was considered day 0 of pseudopregnancy. The selection of days of pseudopregnancy for study was based upon previous observations that progesterone secretion increases after administration of hCG to reach a maximum on day 11 and then declines to day 1 rates by day 18 (Dharmarajan *et al.*, 1989).

Treatment groups

Experiment 1: natural luteal regression. Sixteen rabbits that received hCG on day 0 to render them pseudopregnant were studied. Laparotomy was performed on days 4, 11, 15 and 20 of pseudopregnancy to obtain specimens of ovarian venous blood and luteal tissue.

Experiment 2: luteal regression induced by oestradiol deprivation. Eighteen animals were randomly assigned to one of six experimental groups (three per treatment group). On day 0 of pseudopregnancy, each rabbit had a single Silastic capsule (Dow-Corning Corp., Midland, MI, USA; 3 cm long, 3.35 mm i.d.) implanted s.c. beneath the base of the neck, which contained oestradiol (2 cm filled length; Sigma Chemicals, St Louis, MO, USA) or was empty (control rabbits) (Dharmarajan *et al.*, 1991). On the tenth day of pseudopregnancy and after continuous subcutaneous administration of oestradiol, group 1 rabbits were subjected to laparotomy and blood and luteal tissue samples were collected. At the same stage of pseudopregnancy (day 10), the capsules in groups 2 and 3 were removed to initiate deprivation of oestradiol and the animals were subjected to laparotomy 12 h (group 2) and 24 h (group 3) thereafter. Animals in group 4 had the capsules removed on day 9 and laparotomy performed 72 h later (day 12). Group 5 rabbits had the oestradiol-filled capsules removed on day 9 of pseudopregnancy and reimplanted 24 h later, then underwent laparotomy on day 12 (48 h after capsule replacement). Control animals had the empty Silastic capsule removed on day 10 and laparotomy performed on day 11.

Sample collection

The animals were anaesthetized with intravenous sodium pentobarbital (32 mg kg⁻¹), given heparin sodium (120 U kg⁻¹) for anticoagulation and then subjected to laparotomy. The anastomotic vessels to the ovaries were ligated with 4-0 silk suture. Each ovarian vein was isolated for 5 mm. A small incision was made in the vessel to allow

cannulation with bevelled polyethylene tubing (NewAge Industries, Willow Grove, PA, USA). Ovarian vein blood samples were obtained for determination of plasma progesterone, prostaglandin F metabolite (PGFM) and, in Expt 2, oestradiol. In addition, animals in Expt 2 had blood samples drawn from the marginal ear vein for measurement of peripheral oestradiol. After collection of the blood, the ovaries were excised. The corpora lutea were dissected free from the stroma and weighed individually. All tissues were frozen and stored at -70°C for subsequent determinations of Mn SOD and lipid peroxide activity.

Superoxide dismutase activity

Manganese SOD (Mn SOD) activity in corpora lutea homogenates was measured by the inhibition of nitroblue tetrazolium (NBT) reduction by xanthine oxidase (Oberley & Spitz, 1984). Tissues were homogenized with a Teflon-Potter homogenizer in ice-cold potassium phosphate buffer (0.05 mol l^{-1} , w/v), pH 7.8, with 15–20 strokes. Xanthine (0.1 mmol l^{-1} ; Sigma Chemical Co., St Louis, MO, USA) and xanthine oxidase (Boehringer Mannheim Biochemicals, Indianapolis, USA), adjusted to cause reduction of NBT at $0.015\text{--}0.025\text{ U mg}^{-1}\text{ protein min}^{-1}$ at 560 nm, were used to generate superoxide anion ($\text{O}_2^{\cdot -}$). NBT ($56\text{ }\mu\text{mol l}^{-1}$; Sigma Chemical Co., St Louis, MO, USA) was added to compete with SOD for $\text{O}_2^{\cdot -}$. Diethylenetriamine pentaacetic acid (1 mmol l^{-1} ; Mallinckrodt, Inc., Paris, KY, USA) was incorporated as metal chelator, and catalase (1.0 unit ; Sigma Chemical Co., St Louis, MO, USA) was used to remove hydrogen peroxide (H_2O_2). Sodium cyanide (5 mmol l^{-1}) was added to inhibit copper-zinc SOD (Oberley & Spitz, 1984). These reagents, contained in a 0.8 ml solution, were added to 0.1 ml of the tissue homogenate. NBT reduction was measured by the change in absorbance at 560 nm. One unit of activity was defined as the amount that inhibited the rate of NBT reduction by 50%. The values were standardized to units $\text{mg}^{-1}\text{ protein}$.

Protein concentration in homogenates was determined by the method of Lowry *et al.* (1951) using a commercial kit (Sigma Chemical Co.).

Lipid peroxide

Lipid peroxidation in corpora lutea was assessed by the thiobarbituric (TBA) reaction, which measures the secondary product of peroxidation, malondialdehyde (MDA) (Ohkawa *et al.*, 1979). Corpora lutea were homogenized with a Teflon-Potter homogenizer in ice-cold 1.15% KCl with 15–20 strokes. To 0.1 ml of a 10% (w/v) tissue homogenate was added 0.2 ml of 8.1% sodium dodecyl sulfate, 1.5 ml of 20% acetic acid solution adjusted to pH 3.5 with NaOH and 1.5 ml of 0.8% aqueous solution of TBA (Sigma Chemical Co., St Louis, MO, USA). The mixture was heated in boiling water for 60 min. After cooling, 1.0 ml of distilled water and 5.0 ml of a mixture of *n*-butanol and pyridine ($15:1\text{ v/v}$) were added and the mixture was vigorously shaken. After centrifugation at 5000 g for 10 min, the organic upper layer was removed and its absorbance at 532 nm was measured. 1,1,3,3-Tetramethoxypropane was used as an external standard, and the amount of lipid peroxidation was expressed as $\text{nmol MDA g}^{-1}\text{ wet weight}$.

Radioimmunoassays for progesterone, prostaglandin and oestradiol

Concentrations of ovarian vein plasma progesterone and oestradiol were measured using a solid-phase kit (Diagnostic Products Co., Los Angeles, CA, USA) in which the steroid antibody is covalently bound to the inner surface of polypropylene assay tubes (Dharmarajan *et al.*, 1989). All samples and standards ($100\text{ }\mu\text{l}$) were assayed in duplicate. The sensitivity was 0.05 ng ml^{-1} and the intra- and interassay variations were 7.5 and 6.6%, respectively, for progesterone and were 6.8 and 4.4%, respectively, for oestradiol. Concentration of prostaglandin F metabolite (PGFM) was measured by methods previously described (Dubin *et al.*, 1979). The intraassay coefficient of variation was $<9\%$ and samples from each day were assayed together to preclude error due to interassay variation.

Statistical analysis

Data were evaluated by analysis of variance with repeated measures followed by Duncan's multiple-range test. Two-group comparisons were made by Student's *t* test. A probability of <0.05 was considered significant.

Results

Experiment 1: natural luteal regression

Ovarian venous plasma progesterone and PGFM. Progesterone concentration in ovarian venous plasma increased from day 4 ($492 \pm 150\text{ ng ml}^{-1}$) to day 11 (1107 ± 135) and then declined to very low values by day 20 (228 ± 64) (Fig. 1a). The concentration of PGFM in ovarian venous plasma was similar throughout the course of pseudopregnancy (Fig. 1b) and was not correlated with progesterone concentration.

Fig. 1. Concentration of (a) ovarian venous progesterone and (b) prostaglandin F metabolite (PGFM), at days 4, 11, 15 and 20 of pseudopregnancy in rabbits (Expt 1). Values with different superscripts were significantly different ($P < 0.01$). There was no statistical difference among PGFM concentrations.

Activity of Mn SOD. Activity of Mn SOD increased significantly from day 4 (96 ± 3 U mg⁻¹ protein) to day 11 (178 ± 39), but then declined significantly by day 20 (34 ± 11) (Fig. 2). The Mn SOD activity was significantly associated with progesterone production by the ovary ($r = 0.687$; $P < 0.01$), but was not correlated with PGFM concentration.

Fig. 2. Mean activity of manganese superoxide dismutase (Mn SOD) in corpora lutea on days 4, 11, 15 and 20 of pseudopregnancy in rabbits (Expt 1). Values with different superscripts differed significantly ($P < 0.05$).

Lipid peroxide. There was no consistent pattern of change in MDA concentration during the course of pseudopregnancy. The concentration on day 4 (73 ± 4 MDA nmol mg⁻¹ protein) was significantly ($P < 0.01$) higher than on day 11 (60 ± 3), day 15 (53 ± 4) and day 20 (60 ± 3). Although the concentration of lipid peroxide on day 15, when Mn SOD activity was highest, was

lower than on any of the other days of pseudopregnancy, there was no significant correlation between lipid peroxide and Mn SOD activity. There was no correlation between lipid peroxide and PGFM concentrations.

Experiment 2: luteal regression induced by oestradiol deprivation

Endogenous oestradiol production. Oestradiol implants significantly suppressed endogenous oestradiol production during mid-pseudopregnancy. Ovarian venous oestradiol concentrations ranged from 295.0 ± 39.0 pg ml⁻¹ in control (sham implant) animals to 37.0 ± 3.6 pg ml⁻¹ in animals with intact oestradiol capsules at the time of laparotomy (group 1). The associated peripheral plasma concentrations of oestradiol were 17.0 ± 5.0 pg ml⁻¹ in control and 33.6 ± 5.3 pg ml⁻¹ in group 1 animals. When oestradiol capsules were removed at 12, 24 or 72 h before laparotomy (groups 2, 3 and 4) or were removed and then reimplanted (group 5), oestradiol concentrations in ovarian venous plasma were significantly higher than those of animals with intact capsules (group 1) ($P < 0.05$) (Fig. 3). Corresponding peripheral concentrations of oestradiol were 18.7 ± 2.6 , 14.0 ± 1.7 , 11.6 ± 0.9 and 22.6 ± 1.1 pg ml⁻¹ for groups 2, 3, 4 and 5, respectively.

Fig. 3. Concentration of oestradiol in ovarian venous plasma of subcutaneous oestradiol-treated and control rabbits (Expt 2). Reduction was more pronounced ($P < 0.01$) when the capsules were in place at the time of blood sampling. Values with different superscripts were significantly different ($P < 0.01$).

Progesterone production. There was no difference in ovarian venous progesterone concentrations on day 10 of pseudopregnancy between control and oestradiol-treated, capsule-intact animals (group 1). Progesterone concentrations had significantly decreased when ovarian vein blood was collected 12 (group 2), 24 (group 3) or 72 h (group 4) after capsule removal ($P < 0.001$). However, when exogenous oestradiol support was reinstated for 48 h having been withdrawn for 24 h (group 5), progesterone concentrations approximated those measured in animals that had not been deprived of oestradiol (Fig. 4a).

Ovarian venous prostaglandin F_{2a} metabolite (PGFM). In contrast to its influence on progesterone production, oestradiol capsule placement, withdrawal with or without reimplantation had no effect on PGFM concentrations (Fig. 4b). There was no correlation between concentrations of progesterone and PGFM among the various subgroups. These findings were similar to those obtained in corpora lutea undergoing natural regression.

Activity of Mn SOD. Activity of Mn SOD was low in control animals on day 10 of pseudopregnancy (Fig. 5). The highest Mn SOD activity was in the corpora lutea of animals that had received

Fig. 4. Concentration of (a) ovarian venous progesterone and (b) prostaglandin F metabolite (PGFM), during luteal regression induced by oestradiol deprivation in rabbits (Expt 2). Values with different superscripts differed significantly ($P < 0.001$).

oestradiol capsule treatment throughout the course of pseudopregnancy (group 1). Withdrawal of exogenous oestradiol support before laparotomy resulted in serial decreases in Mn SOD activity at 12, 24 and 72 h after capsule removal. Mn SOD activity increased significantly after restoration of oestradiol capsule treatment in animals that had been deprived of exogenous oestradiol for 24 h during mid-pseudopregnancy (group 5, $P < 0.01$).

Lipid peroxide. Oestradiol capsule implantation on day 0 of pseudopregnancy or capsule removal on day 9 or 10 did not affect lipid peroxide concentrations unless the animals were deprived of exogenous oestradiol for 72 h (Fig. 6), when they significantly increased ($P < 0.01$). Reimplantation of the capsule for 48 h after 24 h of oestradiol deprivation resulted in higher lipid peroxide concentrations than those measured 72 h after withdrawal of exogenous oestradiol ($P < 0.01$).

Discussion

Reactive oxygen species may induce tissue injury through several mechanisms. Although O_2^- is charged, it can cross plasma membranes via anion channels, directly permeate defined lipid bilayers or gain access to intracellular sites as the uncharged $HO_2^\bullet$ (Weiss, 1986). Hydrogen peroxide is a small, neutral molecule that has a permeability constant across membranes that is comparable to that of water (Chance *et al.*, 1979). An oxidant attack on membrane, lipoprotein or albumin-

Fig. 5. Activity of manganese superoxide dismutase (Mn SOD) in corpora lutea (Expt 2) of rabbits. Activity was low in control, increased to maximum with continuous oestradiol treatment and decreased significantly after capsule removal ($P < 0.01$). Means within treatment group with different letter designations were significantly different ($P < 0.01$).

Fig. 6. Lipid peroxidation in corpora lutea of rabbits as assessed by malondialdehyde (MDA) concentration (Expt 2). Oestradiol capsule implantation or removal did not affect lipid peroxide levels 72 h after removal, at which time the level was significantly increased ($P < 0.01$). Reimplantation further increased lipid peroxide over that measured after 72 h of deprivation of exogenous oestradiol ($P < 0.01$). Values with different superscripts were significantly different ($P < 0.01$).

associated polyunsaturated fatty acids can initiate a complex cascade of events leading to the formation of reactive, unstable oxidants, long-lived toxic by-products, or biologically active inflammatory mediators that have the potential to propagate damage beyond the confines of the original focus. Furthermore, superoxides may directly alter protein and nucleic acid structure or function (Freeman & Crapo, 1982).

In addition to the antioxidant compounds of tocopherol and ascorbate, the ovary contains SOD, catalase and glutathione peroxidase which enzymatically inactivate these highly toxic oxygen metabolites. Superoxide dismutase, the specific inhibitor of superoxide anion radical, has been isolated in several forms, which differ in their transition metal at the active centre and in their cell

location. Copper–zinc SOD is primarily found in the cytosol and extracellular space, whereas Mn SOD is isolated in the mitochondrial matrix (Fridovich, 1986). The latter metalloenzyme is of particular relevance to corpus luteum function, for oxygen free radicals are produced not only by leucocytes but also by the mitochondrial electron-transport system and cytochrome P450_{scc} (Freeman & Crapo, 1982; Weiss, 1986). Since cytochrome P450_{scc} is a key enzyme in progesterone synthesis, the reactive oxygen species created during the process of steroidogenesis may cause local damage and decrease steroidogenesis if not detoxified by SOD and other scavengers.

The presence of superoxide anions can induce both Cu–Zn SOD and Mn SOD (Dryer *et al.*, 1980), although *in vitro* studies have demonstrated that, in very high concentrations, H₂O₂ or a combination of O₂^{•-} and H₂O₂ may inhibit the activity of SOD (Sinet & Garber, 1981). In addition, the toxic hydroxyl radical may be produced by the Haber–Weiss reaction when H₂O₂, the end-product of SOD, reacts with O₂^{•-} in the presence of iron (Fridovich, 1986). Without the presence of sufficient activity of the cytosolic enzyme, catalase, H₂O₂ generated in the dismutation reaction could itself inhibit progesterone production (Riley & Behrman, 1991). Recent data suggest that H₂O₂ may affect progesterone synthesis by blocking the transfer of intracellular cholesterol to mitochondria or the translocation of cholesterol across the outer mitochondrial membrane (Behrman & Aten, 1991).

The results of this study demonstrate the existence of a dependent relationship between Mn SOD activity and progesterone secretion in both natural luteolysis and regression induced by oestradiol deprivation. Concentrations of Mn SOD and progesterone production were high on day 11 of pseudopregnancy in the animals studied in Expt 1. The significant decline in Mn SOD activity on days 15 and 20 was matched by a fall in progesterone production by the ovary. In the capsule model, oestradiol deprivation of the corpus luteum for 12 or more hours at mid-pseudopregnancy resulted in the expected fall in progesterone secretion (Dharmarajan *et al.*, 1991), but also a precipitous drop in measured SOD. This association between SOD and corpus luteum function is consistent with the hypothesis that very high concentrations of oxygen free radicals may be present at the end of the life span of the corpus luteum which inactivate the cellular defences provided by SOD, promote tissue damage and further reduce the availability of protective enzymes such as SOD. Replacement of the oestradiol capsule after temporary withdrawal of exogenous hormonal support led to a rise in Mn SOD activity and reversal of functional regression, which gives further credence to the proposed relationships described.

Very low, yet stable, concentrations of progesterone were detected in the ovarian vein at 12, 24 and 72 h after removal of the oestradiol capsule, despite a continued decline in Mn SOD activity. The corpus luteum may have a reserve capacity for progesterone synthesis during this period of structural regression. Recent data from our laboratory have demonstrated that, after unilateral luteectomy or oophorectomy in pseudopregnant rabbits, ovarian venous progesterone concentration and progesterone secretion rates in the intact ovary increased to nearly 70% above their normal values (Dharmarajan *et al.*, 1992). Hence, given the appropriate stimulus, luteal tissue may react to alterations in the hormonal milieu by mechanisms that attempt to sustain steroid secretion.

One process by which oestradiol may regulate luteal steroidogenic function in rabbits is through direct or indirect stimulation of free-radical scavengers. Continuous subcutaneous administration of oestradiol during early and mid-pseudopregnancy yielded a supraphysiological concentration of circulating plasma oestradiol and suppressed ovarian follicular production of oestradiol. The corpus luteum responded to this high oestradiol stimulation with a significant increase in luteal SOD activity, compared with control animals. The decline in exogenous circulating oestradiol after capsule removal led to partial recovery of ovarian follicular oestradiol production, but the overall diminution in oestradiol stimulation of the corpus luteum that occurred was associated with a fall in SOD activity over time. With restoration of oestradiol support after 24 h of depletion, SOD activity again rose, suggesting a dependent relationship.

In contrast to the data supporting a role for reactive oxygen species in functional luteolysis of pseudopregnant rabbits, the lack of significant change in prostaglandin metabolites during natural

regression or that induced by oestradiol depletion suggests that prostaglandins do not play a primary role in luteolysis in rabbits. This corroborates the results of an earlier study where prostaglandin concentrations were measured in the tissues and *in vitro* perfusates of ovaries in late pseudopregnancy (Dharmarajan *et al.*, 1989). The lack of correlation between the activity of the free-radical scavenger and prostaglandin metabolites would have been impossible to demonstrate if luteolysis had been initiated by prostaglandin administration rather than oestradiol deprivation. The use of prostaglandins may have created conditions that are not necessarily characteristic of natural luteolysis, for complex interactions exist among reactive oxygen species and the products of the lipoxygenase and cyclooxygenase pathways (Hemler *et al.*, 1979; Parente, 1982).

An increased production of lipid peroxide was not noted in corpora lutea obtained during late pseudopregnancy, but, in the oestradiol deprivation model, luteal tissue lipid peroxide increased if the steroid capsule had been removed 72 h before sampling. This rise was not seen 24 h after capsule removal, although the decline in progesterone secretion had occurred by this time. Previous data concerning natural and prostaglandin-induced luteal regression in rats revealed that lipid peroxides increased after the initial decline in progesterone secretion was measured (Sawada & Carlson, 1985, 1991). This temporal relationship suggests that lipid peroxidation is not responsible for initiating the functional changes associated with luteolysis, although membrane breakdown is an important feature of regression.

High concentrations of lipid peroxide were noted in animals of group 5 that experienced oestradiol deprivation followed by restoration of exogenous oestradiol support for 48 h before collection of corpora lutea. The reason is not readily apparent, but may be associated with an impaired ability of the luteal cell that had been previously damaged by oestradiol deprivation to detoxify reactive oxygen species. The heightened metabolic state of rabbit mitochondria that occurred with replacement of the major luteotrophic hormone, oestradiol, caused a proportional increase in production of O_2^- and H_2O_2 (Dryer *et al.*, 1980). Nevertheless, the activity of Mn SOD did not approximate to that measured in animals receiving continuous oestradiol support and may reflect insufficient scavenging ability to prevent lipid oxidation. This hypothesis requires further investigation.

The results from the present study demonstrate a relationship between oestradiol concentration, Mn SOD activity and progesterone production in corpora lutea of pseudopregnant rabbits undergoing natural luteolysis and demise induced by depletion of oestradiol. A decline in antioxidant capability of the regressing corpus luteum may result in increased local concentrations of oxygen free radicals, which stimulate functional and structural regression. If so, it is achieved through mechanisms that do not result in heightened production of prostaglandin F metabolites. The association between changes in luteotrophic stimulation by oestradiol and Mn SOD activity suggests that oestradiol may regulate corpus luteum steroidogenesis through this superoxide scavenger, which is primarily localized to the mitochondrial matrix of cells.

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References

- Behrman, H.R. & Aten, R.F. (1991) Evidence that hydrogen peroxide blocks hormone-sensitive cholesterol transport into mitochondria of rat luteal cells. *Endocrinology* **128**, 2958–2966.
- Behrman, H.R. & Preston, S.L. (1989) Luteolytic actions of peroxide in rat ovarian cells. *Endocrinology* **124**, 2895–2900.
- Bill, II, C.H. & Keyes, P.L. (1983) 17β -estradiol maintains normal function of corpora lutea throughout pseudo-pregnancy in hypophysectomized rabbits. *Biology of Reproduction* **28**, 608–617.
- Chance, B., Sies, H. & Boveris, A. (1979) Hydroperoxide metabolism in mammalian organs. *Physiological Reviews* **59**, 527–605.

- Dharmarajan, A.M., Sueoka, K., Miyazaki, T., Atlas, S.J., Ghodgaonkar, R.B., Dubin, N.H., Zirkin, B.R. & Wallach, E.E. (1989) Prostaglandins and progesterone secretion in the *in vitro* perfused pseudopregnant rabbit ovary. *Endocrinology* **124**, 1198–1203.
- Dharmarajan, A.M., Zanagnolo, V.L., Dasko, L.M., Zirkin, B.R., Ewing, L.L. & Wallach, E.E. (1991) Estradiol regulation of the rabbit corpus luteum: *in vivo* and *in vitro* studies. *Endocrinology* **128**, 2678–2684.
- Dharmarajan, A.M., Zanagnolo, V.L., Dasko, L.M., Hardy, M.P. & Wallach, E.E. (1992) Changes in rabbit corpus luteum progesterone secretion and cellular morphology following unilateral luteectomy or ovariectomy. *Biology of Reproduction* **46**, 251–255.
- Dryer, S.E., Dryer, R.L. & Autor, A.P. (1980) Enhancement of mitochondrial, cyanide-resistant superoxide dismutase in the livers of rats treated with 2,4-dinitrophenol. *Journal of Biological Chemistry* **255**, 1054–1057.
- Dubin, N.H., Ghodgaonkar, R.B. & King, T.M. (1979) Role of prostaglandin production in spontaneous and oxytocin-induced uterine contractile activity in *in vitro* pregnant rat uteri. *Endocrinology* **105**, 47–51.
- Freeman, B.A. & Crapo, J.D. (1982) Free radicals and tissue injury. *Laboratory Investigation* **47**, 412–426.
- Fridovich, I. (1986) Biological effects of the superoxide radical. *Archives of Biochemistry and Biophysics* **247**, 1–11.
- Gatzuli, E., Aten, R.F. & Behrman, H.R. (1991) Inhibition of gonadotropin action and progesterone synthesis by xanthine oxidase in rat luteal cells. *Endocrinology* **128**, 2253–2258.
- Hemler, M.D., Cook, H.W. & Lands, W.E. (1979) Prostaglandin biosynthesis can be triggered by lipid peroxides. *Archives of Biochemistry and Biophysics* **193**, 340–345.
- Holt, J.A. (1989) Regulation of progesterone production in the rabbit corpus luteum. *Biology of Reproduction* **40**, 201–208.
- Laloraya, M., Kumar, G.P. & Laloraya, M.M. (1988) Changes in the levels of superoxide anion radical and superoxide dismutase during the estrous cycle of *Rattus norvegicus* and induction of superoxide dismutase in rat ovary by lutropin. *Biochemical and Biophysical Research Communications* **157**, 146–153.
- Lowry, O.H., Rosenbrough, N.J., Farr, A.L. & Randall, R.J. (1951) Protein measurement with the Folin phenol reagent. *Journal of Biological Chemistry* **193**, 265–275.
- Miyazaki, T., Sueoka, K., Dharmarajan, A.M., Atlas, S.J., Bulkley, G.B. & Wallach, E.E. (1991) Effect of inhibition of oxygen free radical on ovulation and progesterone production by the *in-vitro* perfused rabbit ovary. *Journal of Reproduction and Fertility* **91**, 207–212.
- Oberley, L.W. & Spitz, D.R. (1984) Assay of superoxide dismutase activity in tumor tissue. *Methods in Enzymology* **105**, 457–464.
- Ohkawa, H., Ohishi, N. & Yagi, K. (1979) Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical Biochemistry* **95**, 351–358.
- Parente, L. (1982) Study on the effect of superoxide dismutase on arachidonic acid metabolism. *Prostaglandins* **23**, 725–730.
- Riley, J.C.M. & Behrman, H.R. (1991) *In vivo* generation of hydrogen peroxide in the rat corpus luteum during luteolysis. *Endocrinology* **128**, 1749–1753.
- Sawada, M. & Carlson, J.C. (1985) Association of lipid peroxidation during luteal regression in the rat and natural aging in the rotifer. *Experimental Gerontology* **20**, 179–186.
- Sawada, M. & Carlson, J.C. (1989) Superoxide radical production in plasma membrane samples from regressing rat corpora lutea. *Canadian Journal of Physiology and Pharmacology* **67**, 465–471.
- Sawada, M. & Carlson, J.D. (1991) Rapid plasma membrane changes in superoxide radical formation, fluidity, and phospholipase A₂ activity in the corpus luteum of the rat during the induction of luteolysis. *Endocrinology* **128**, 2992–2997.
- Sinet, P.-M. & Garber, P. (1981) Inactivation of the human CuZn superoxide dismutase during exposure to O₂⁻ and H₂O₂. *Archives of Biochemistry and Biophysics* **212**, 411–416.
- Weiss, S.J. (1986) Oxygen, ischemia and inflammation. *Acta Physiologica Scandinavica Supplementum* **548**, 9–37.

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